

Untitled

J Lacasa

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Libraries

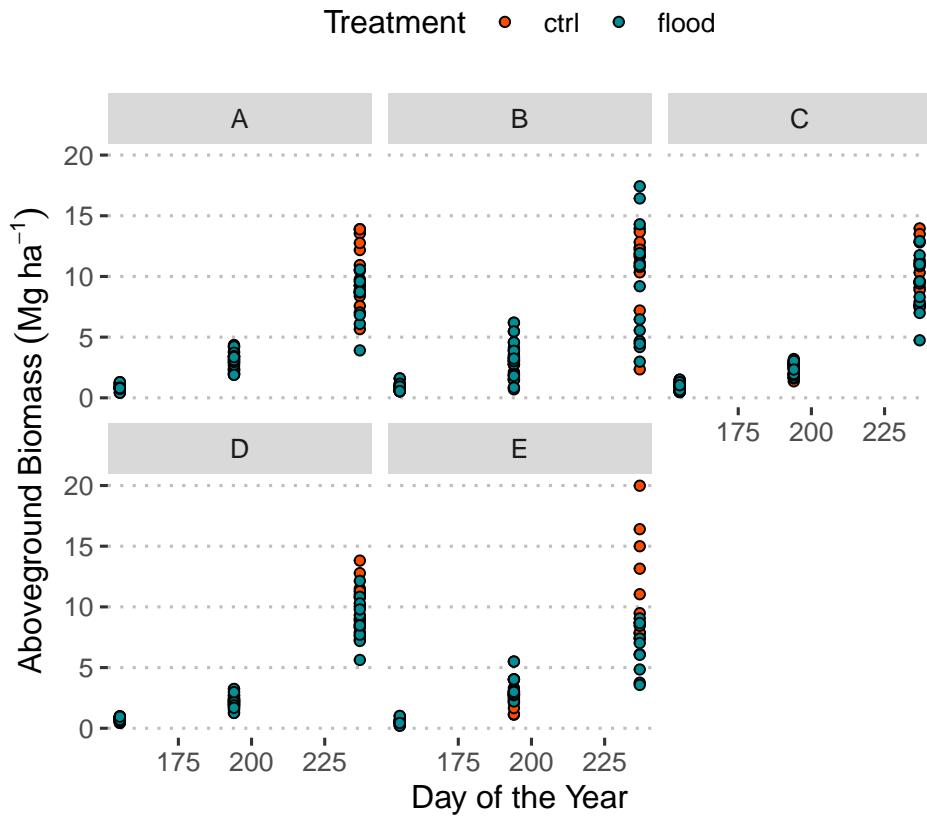
```
library(tidyverse) # data wrangling, data viz
library(mgcv) # fitting generalized additive models
library(latex2exp) # math notation in plots
library(ggpubr) # plot aesthetics
```

Loading the data

```
# dd_finalproj <- read.csv("../classes/data/dd_finalproj.csv")
url <- "https://raw.githubusercontent.com/jlacasa/stat705_fall2024/main/classes/data/dd_finalproj.csv"
dd_finalproj <- read.csv(url)
dd_finalproj$doy_f <- factor(dd_finalproj$doy)
```

Exploratory Data Analysis

```
dd_finalproj %>%
  ggplot(aes(doy, fitted))+
  geom_point(aes(y = agb_g, fill = trt), shape = 21)+
  facet_wrap(~species)+
  scale_fill_manual(values = c("#FC4C02", "#008E97"))+
  labs(y = expression(Aboveground~Biomass~(Mg~ha^{-1})),
       x= "Day of the Year",
       color = "Treatment",
       fill = "Treatment")+
  theme_pubclean()+
  theme(aspect.ratio = 1)
```



Model Fitting

Firstly, fit a simple model

$$y_{ijkl} \sim N(\mu_{ijk}, \sigma^2)$$

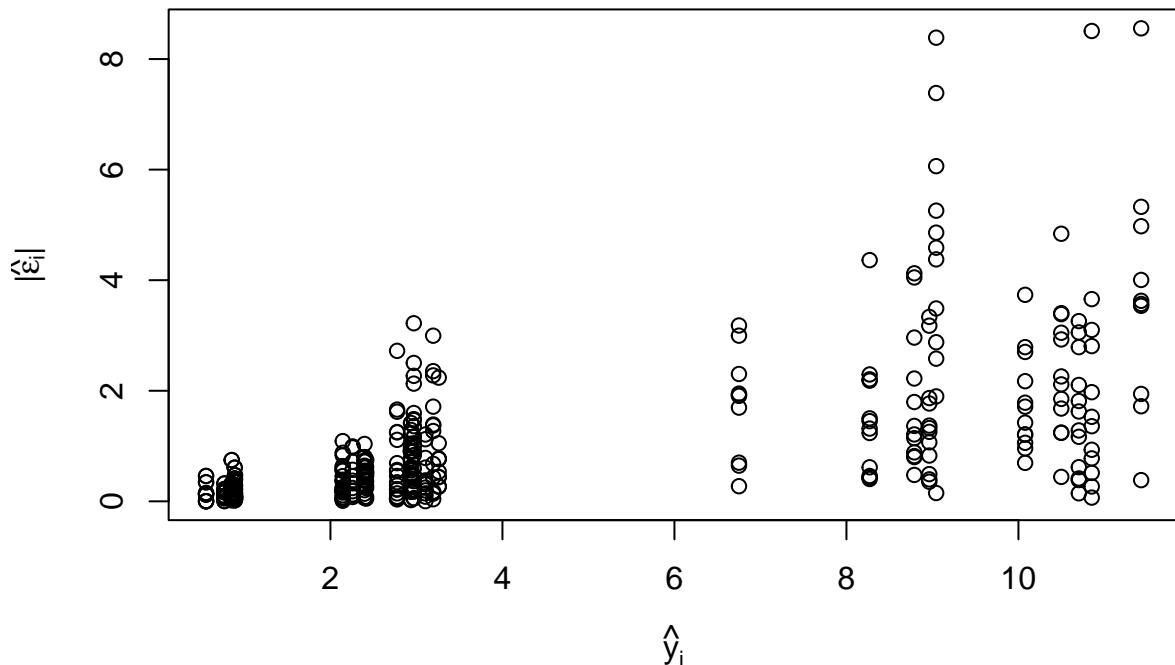
$$\mu_{ijk} = \beta_0 + \tau_i + \rho_j + \alpha_k + (\tau\rho)_{ij} + (\tau\alpha)_{ik} + (\rho\alpha)_{ik} + (\tau\rho\alpha)_{ijk},$$

where y_{ijkl} is the observation of aboveground biomass (in g) for the i th treatment, j th species, k th moment (i.e., time), and l th repetition, that arises from a Normal distribution with mean μ_{ijk} and variance σ^2 . The β_0 is the general intercept, τ_i is the effect of the i th treatment, ρ_j is the effect of the j th species, and $(\tau\rho)_{ij}$, $(\tau\alpha)_{ik}$, and $(\rho\alpha)_{ik}$ are the two-way interactions between the factors mentioned above, and $(\tau\rho\alpha)_{ijk}$ is the three-way interaction of the factors mentioned above. Note that there is a single variance for all observations.

Model diagnostics

```
plot(m1$fitted.values, abs(residuals(m1, type = "deviance")),
     xlab = TeX("\$\\hat{y}_i\$"),
     ylab = TeX("|\$\\hat{\epsilon}_i\$|"),
     main = "Residuals versus fitted values",
     sub = "Note that the errors increase together with the means")
```

Residuals versus fitted values



Constant variance

Note that the errors increase together with the means

```
lm(log(agb_g) ~ species*trt*doy_f, data = dd_finalproj)
```

```
##
## Call:
## lm(formula = log(agb_g) ~ species * trt * doy_f, data = dd_finalproj)
##
## Coefficients:
##             (Intercept)          speciesB
##             -1.514e-01         -6.419e-02
##             speciesC           speciesD
##             -3.885e-02         -1.388e-01
##             speciesE           trtflood
##             -5.354e-01          8.716e-15
##             doy_f194            doy_f237
##             1.194e+00           2.468e+00
##             speciesB:trtflood speciesC:trtflood
##             -9.031e-15          -9.126e-15
##             speciesD:trtflood speciesE:trtflood
##             -5.422e-15          -9.659e-15
##             speciesB:doy_f194 speciesC:doy_f194
##             -1.057e-02          -1.565e-01
##             speciesD:doy_f194 speciesE:doy_f194
##             -1.646e-01          4.488e-01
##             speciesB:doy_f237 speciesC:doy_f237
##             5.510e-02           7.766e-02
```

```

##          speciesD:doy_f237           speciesE:doy_f237
##                1.135e-01            5.849e-01
##          trtflood:doy_f194        trtflood:doy_f237
##                6.994e-02            -2.364e-01
## speciesB:trtflood:doy_f194  speciesC:trtflood:doy_f194
##                -2.277e-03            -5.797e-02
## speciesD:trtflood:doy_f194  speciesE:trtflood:doy_f194
##                -2.403e-02            1.197e-01
## speciesB:trtflood:doy_f237  speciesC:trtflood:doy_f237
##                -2.553e-02            2.229e-02
## speciesD:trtflood:doy_f237  speciesE:trtflood:doy_f237
##                1.190e-01            -2.714e-01

```

Model fitting II

The first, simpler, model (i.e., `m1`) does not seem to have constant variance. We can follow two routes: (1) transform the response and keep the assumptions, or (2) keep the data as is and change our model assumptions.

Following the second option, we can model

$$y_{ijkl} \sim N(\mu_{ijk}, \sigma_k^2)$$

$$\mu_{ijk} = \beta_0 + \tau_i + \rho_j + \alpha_k + (\tau\rho)_{ij} + (\tau\alpha)_{ik} + (\rho\alpha)_{ik} + (\tau\rho\alpha)_{ijk},$$

where y_{ijkl} is the observation of aboveground biomass (in g) for the i th treatment, j th species, k th moment (i.e., time), and l th repetition, that arises from a Normal distribution with mean μ_{ijk} and variance at time k , σ_k^2 . The β_0 is the general intercept, τ_i is the effect of the i th treatment, ρ_j is the effect of the j th species, and $(\tau\rho)_{ij}$, $(\tau\alpha)_{ik}$, and $(\rho\alpha)_{ik}$ are the two-way interactions between the factors mentioned above, and $(\tau\rho\alpha)_{ijk}$ is the three-way interaction of the factors mentioned above. Note that the variance is a function of time.

```

m2 <- gam(list(agb_g ~ species*trt*factor(doy),
               ~ doy),
           family = gaulss(),
           data = dd_finalproj)
m2_fitted <- predict(m2)[,1]
m2_residuals <- residuals(m2, type = "deviance")

```

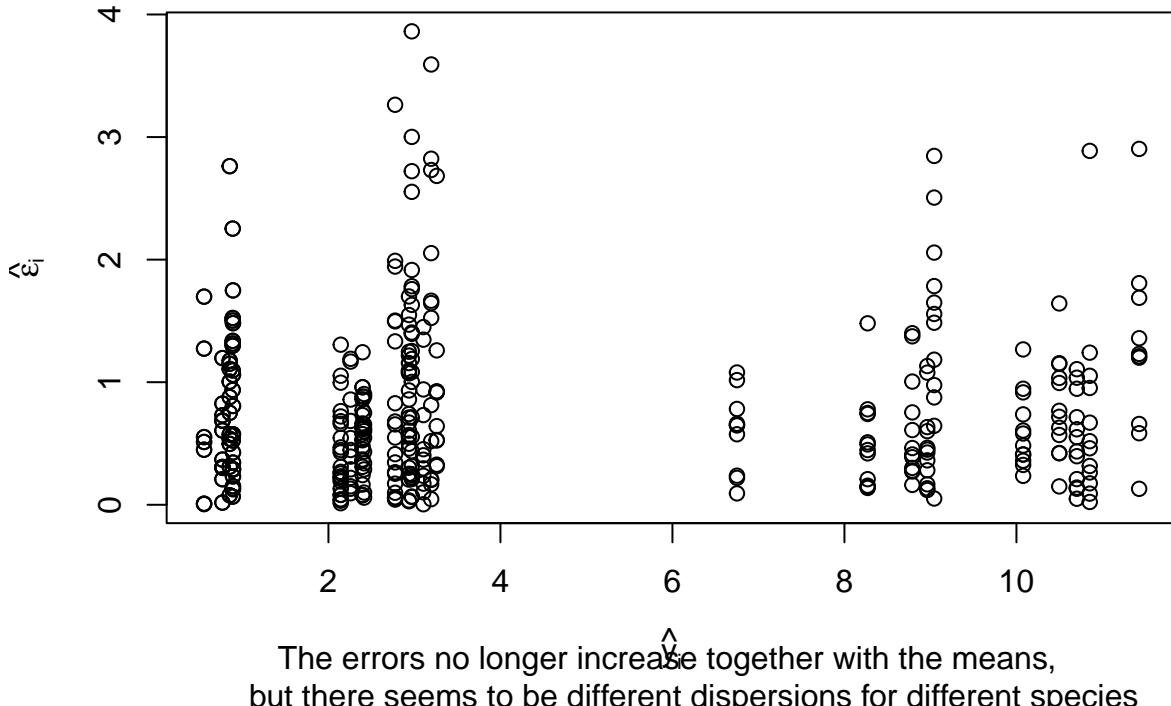
Model diagnostics II

```

plot(m2_fitted, abs(m2_residuals),
      xlab = TeX("$\\hat{y}_i$"),
      ylab = TeX("$\\hat{\\epsilon}_i$"),
      main = "Residuals versus fitted values",
      sub = "The errors no longer increase together with the means,
             but there seems to be different dispersions for different species")

```

Residuals versus fitted values



Model fitting III

The second, model (i.e., m2) fixes the issue of constant variance. However, the variance seems to be different depending on the groups.

We can thus further model σ^2 :

$$y_{ijkl} \sim N(\mu_{ijk}, \sigma_{jk}^2)$$

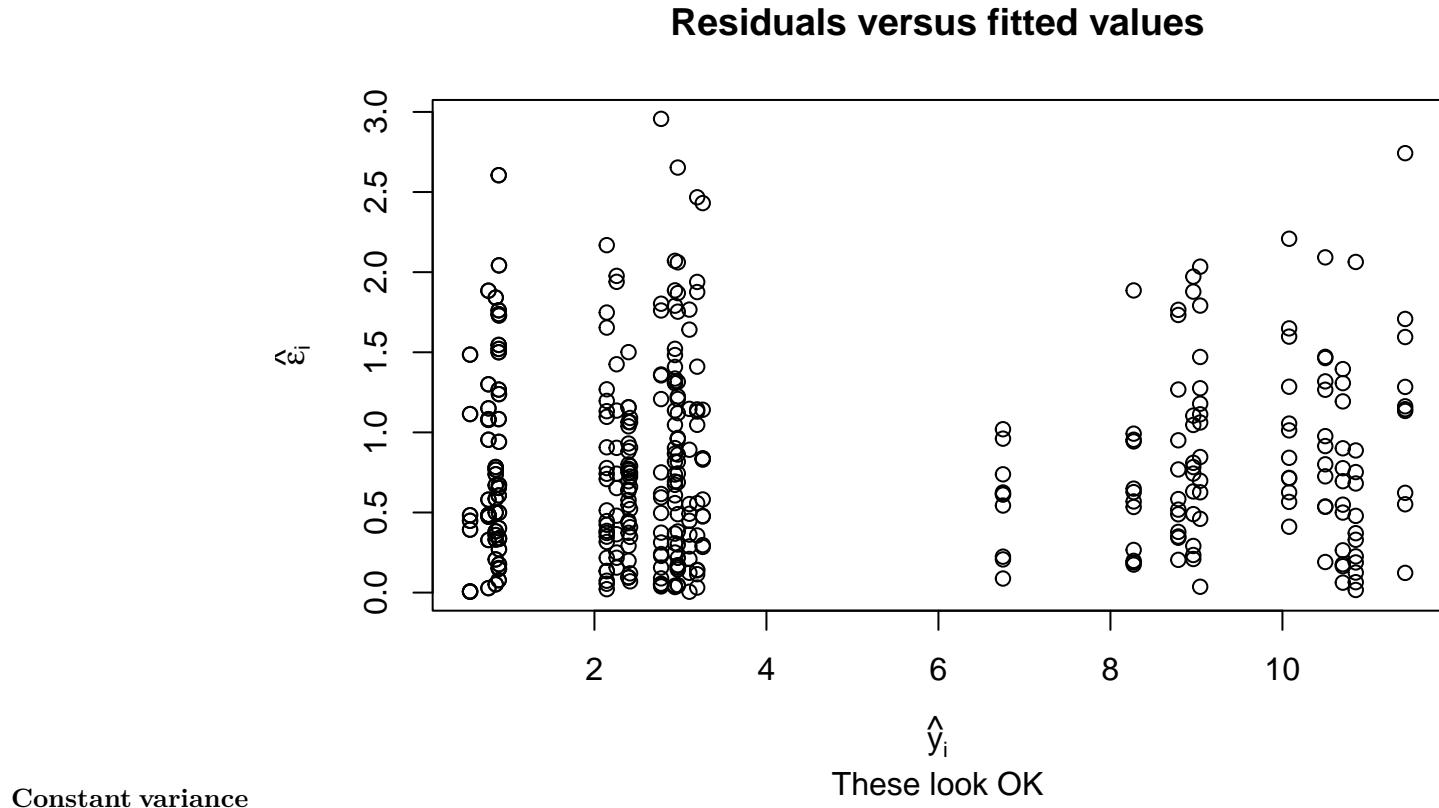
$$\mu_{ijk} = \beta_0 + \tau_i + \rho_j + \alpha_k + (\tau\rho)_{ij} + (\tau\alpha)_{ik} + (\rho\alpha)_{ik} + (\tau\rho\alpha)_{ijk},$$

where y_{ijkl} is the observation of aboveground biomass (in g) for the i th treatment, j th species, k th moment (i.e., time), and l th repetition, that arises from a Normal distribution with mean μ_{ijk} and variance at time k for species j , σ_{jk}^2 . The β_0 is the general intercept, τ_i is the effect of the i th treatment, ρ_j is the effect of the j th species, and $(\tau\rho)_{ij}$, $(\tau\alpha)_{ik}$, and $(\rho\alpha)_{ik}$ are the two-way interactions between the factors mentioned above, and $(\tau\rho\alpha)_{ijk}$ is the three-way interaction of the factors mentioned above. Note that the variance is a function of time.

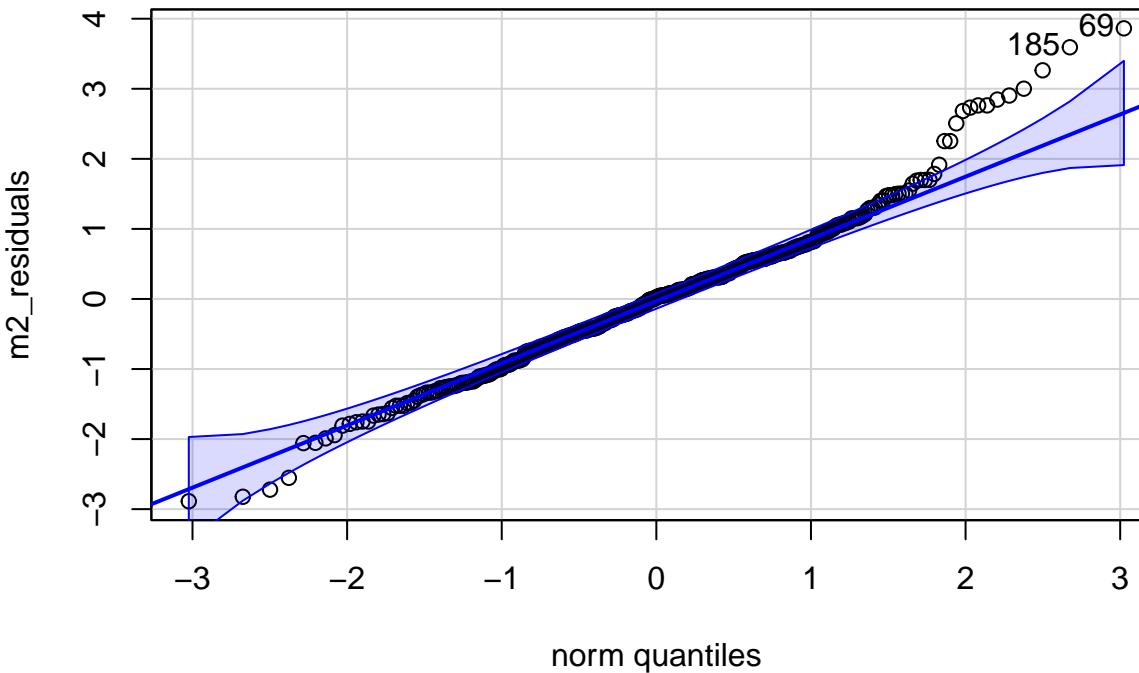
```
m3 <- gam(list(agb_g ~ species*trt*factor(doy),
  ~ doy + species),
  family = gaulss(),
  data = dd_finalproj)
m3_fitted <- predict(m3)[,1]
m3_residuals <- residuals(m3, type = "deviance")
```

Model diagnostics III

```
plot(m3_fitted, abs(m3_residuals),
     xlab = TeX("$\\hat{y}_i$"),
     ylab = TeX("$|\\hat{\\epsilon}_i|$"),
     main = "Residuals versus fitted values",
     sub = "These look OK")
```



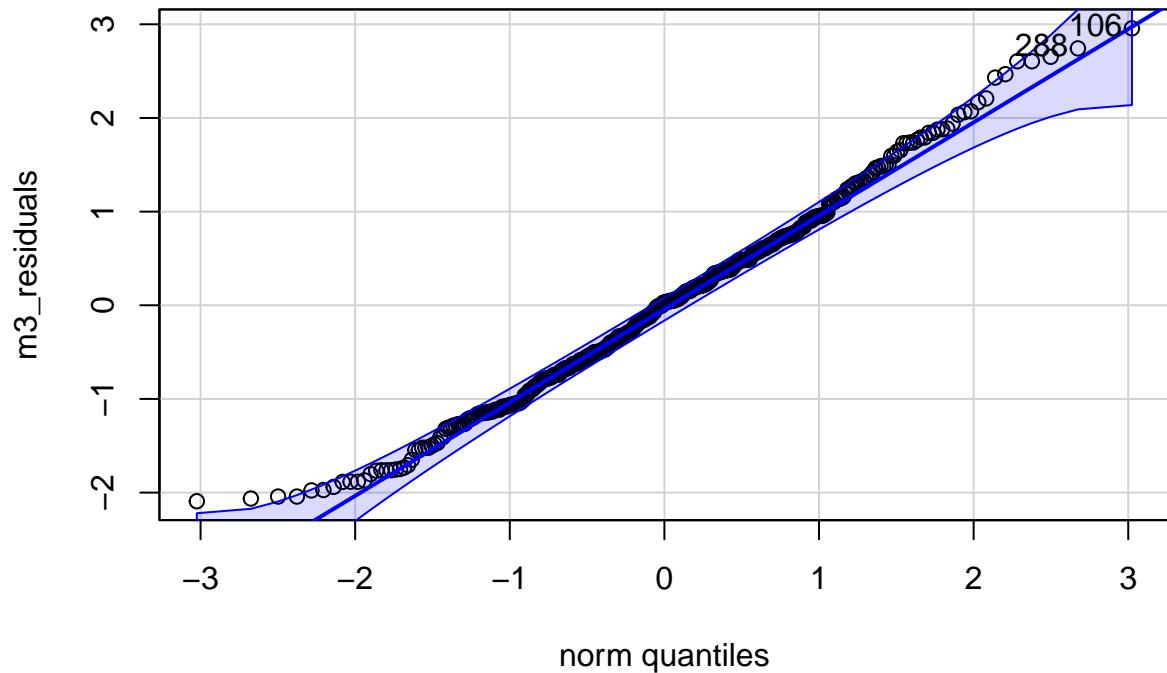
```
car::qqPlot(m2_residuals)
```



Normal distribution

```
## [1] 69 185
```

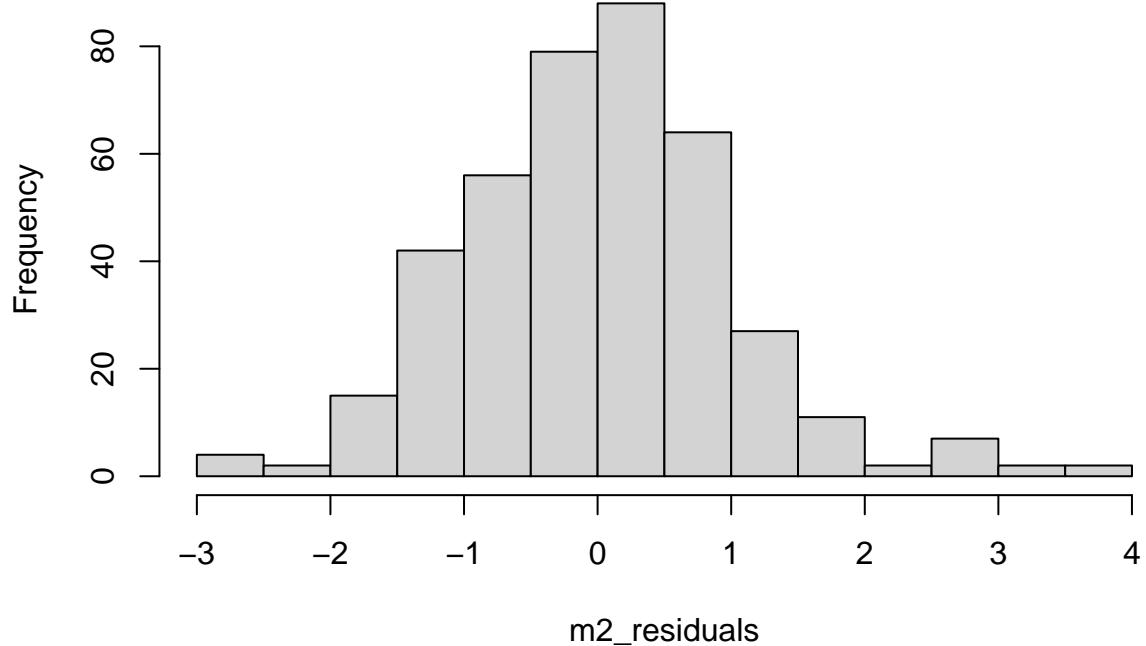
```
car::qqPlot(m3_residuals)
```



```
## [1] 106 288
```

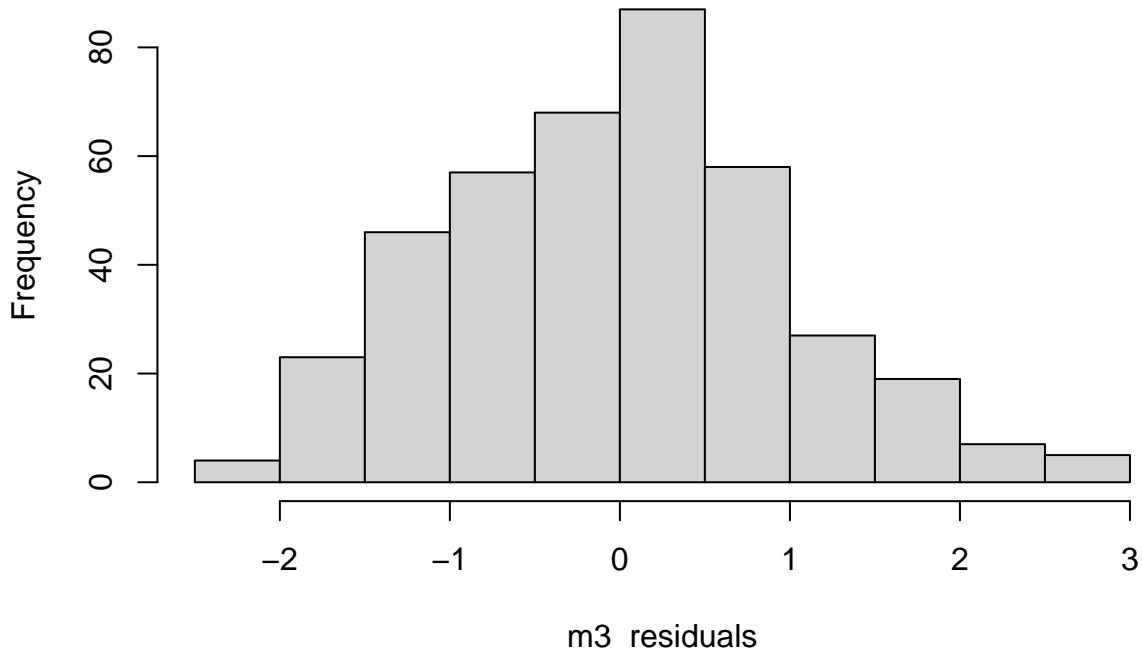
```
hist(m2_residuals)
```

Histogram of m2_residuals



```
hist(m3_residuals)
```

Histogram of m3_residuals



AIC, BIC

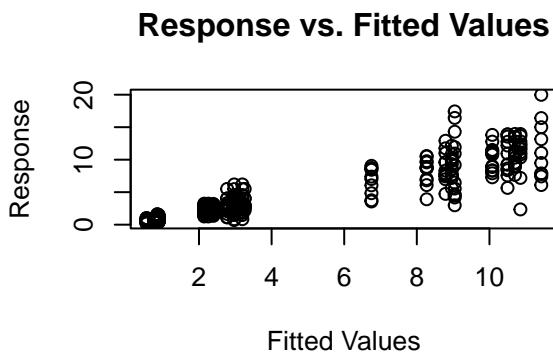
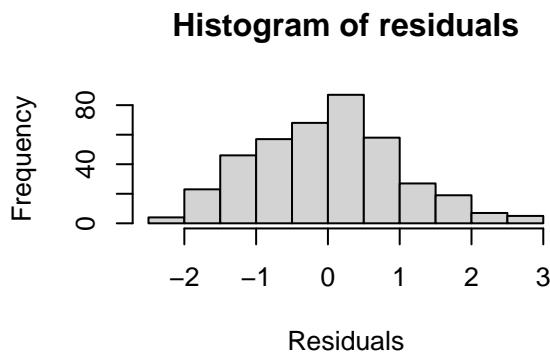
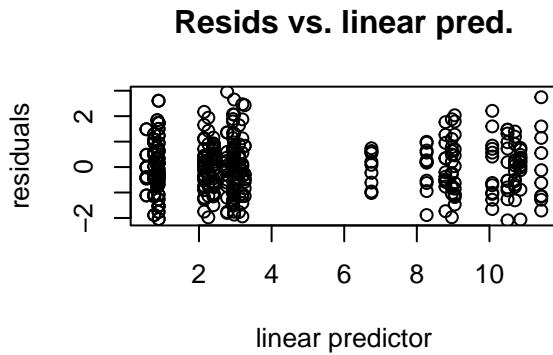
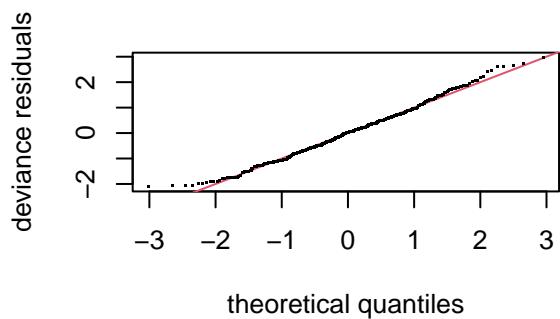
```
left_join(AIC(m1,m2,m3) %>% rownames_to_column("model"),
          BIC(m1,m2,m3) %>% rownames_to_column("model"))

## Joining with 'by = join_by(model, df)'

##   model df      AIC      BIC
## 1     m1 31 1594.928 1718.741
## 2     m2 32 1101.115 1228.922
## 3     m3 36 1034.997 1178.780
```

Other checks

```
gam.check(m3)
```



```
## 
## Method: REML   Optimizer: outer newton
## Model required no smoothing parameter selectionModel rank =  36 / 36
```

Inference

```
summary(m3)
```

Summary

```
## 
## Family: gaulss
## Link function: identity logb
## 
## Formula:
## agb_g ~ species * trt * factor(doy)
## ~doy + species
## 
## Parametric coefficients:
##                               Estimate Std. Error z value Pr(>|z|)
## (Intercept)                8.900e-01  6.681e-02 13.321 < 2e-16 ***
## speciesB                 -3.767e-02  1.347e-01 -0.280 0.779799
```

```

## speciesC          -3.053e-03  9.495e-02 -0.032  0.974351
## speciesD          -1.224e-01  8.317e-02 -1.471  0.141218
## speciesE          -3.361e-01  1.345e-01 -2.498  0.012475 *
## trtflood          1.820e-17  9.448e-02  0.000  1.000000
## factor(doy)194   2.047e+00  1.549e-01 13.215 < 2e-16 ***
## factor(doy)237   9.608e+00  6.711e-01 14.316 < 2e-16 ***
## speciesB:trtflood 4.215e-17  1.906e-01  0.000  1.000000
## speciesC:trtflood -3.364e-18  1.343e-01  0.000  1.000000
## speciesD:trtflood -2.778e-17  1.176e-01  0.000  1.000000
## speciesE:trtflood -4.214e-17  1.902e-01  0.000  1.000000
## speciesB:factor(doy)194 7.061e-02  3.148e-01  0.224  0.822546
## speciesC:factor(doy)194 -5.368e-01  2.201e-01 -2.439  0.014741 *
## speciesD:factor(doy)194 -6.710e-01  1.922e-01 -3.490  0.000483 ***
## speciesE:factor(doy)194 1.749e-01  2.828e-01  0.618  0.536332
## speciesB:factor(doy)237 3.923e-01  1.371e+00  0.286  0.774820
## speciesC:factor(doy)237 2.076e-01  9.540e-01  0.218  0.827742
## speciesD:factor(doy)237 -2.978e-01  8.314e-01 -0.358  0.720210
## speciesE:factor(doy)237 1.265e+00  1.199e+00  1.055  0.291202
## trtflood:factor(doy)194 1.676e-01  2.598e-01  0.645  0.518822
## trtflood:factor(doy)237 -2.228e+00  9.491e-01 -2.347  0.018906 *
## speciesB:trtflood:factor(doy)194 5.764e-02  5.194e-01  0.111  0.911626
## speciesC:trtflood:factor(doy)194 -1.499e-01  3.693e-01 -0.406  0.684691
## speciesD:trtflood:factor(doy)194 -5.262e-02  3.224e-01 -0.163  0.870349
## speciesE:trtflood:factor(doy)194 3.163e-01  4.710e-01  0.672  0.501862
## speciesB:trtflood:factor(doy)237 4.189e-01  1.939e+00  0.216  0.828968
## speciesC:trtflood:factor(doy)237 3.146e-01  1.349e+00  0.233  0.815607
## speciesD:trtflood:factor(doy)237 1.114e+00  1.176e+00  0.947  0.343525
## speciesE:trtflood:factor(doy)237 -2.449e+00  1.695e+00 -1.444  0.148625
## (Intercept).1        -5.935e+00  2.492e-01 -23.814 < 2e-16 ***
## doy.1                  2.856e-02  1.201e-03 23.775 < 2e-16 ***
## speciesB.1              5.797e-01  1.105e-01  5.248  1.53e-07 ***
## speciesC.1              1.026e-02  1.129e-01  0.091  0.927546
## speciesD.1              -3.148e-01  1.117e-01 -2.819  0.004813 **
## speciesE.1              3.001e-01  1.195e-01  2.511  0.012040 *
## ---
## Signif. codes:  0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1
##
##
## Deviance explained = 98.4%
## -REML =  512.5  Scale est. = 1           n = 401

```

```
anova(m3)
```

ANOVA

```

## 
## Family: gaulss
## Link function: identity logb
## 
## Formula:
## agb_g ~ species * trt * factor(doy)

```

```

## ~doy + species
##
## Parametric Terms:
##                               df   Chi.sq  p-value
## species                  4    8.404  0.07785
## trt                      1    0.000  1.00000
## factor(doy)              2  364.014 < 2e-16
## species:trt               4    0.000  1.00000
## species:factor(doy)       8   23.406  0.00288
## trt:factor(doy)           2    6.044  0.04870
## species:trt:factor(doy)   8   6.547  0.58619
## doy.1                     1  565.269 < 2e-16
## species.1                 4   73.347 4.46e-15

```

```

means <- emmeans::emmeans(m3,
                           ~trt:species, at = list("doy" = c(237)))

mean_comparisons <- multcomp::cld(means,
                                      level = 0.05,
                                      adjust = "none",
                                      decreasing = TRUE,
                                      Letters = letters) %>%
  mutate(.group = trimws(.group))

mean_comparisons %>%
  transmute(species, trt, SE=round(SE,2),
            emmean = round(emmean, 2), group = .group,
            lower.CL = round(lower.CL,2), upper.CL = round(upper.CL,2))

```

Means, Mean comparisons

```

##   species   trt   SE emmean group lower.CL upper.CL
## 10      E  ctrl 0.99  12.56     a   12.50   12.62
## 2       B  ctrl 1.19  12.27    ab   12.19   12.34
## 8       C  ctrl 0.68  11.55     a   11.50   11.59
## 6       A  ctrl 0.67  11.33    ab   11.29   11.37
## 4       D  ctrl 0.50  10.60   abc  10.57   10.63
## 7      B flood 1.19  10.46  abcd  10.38   10.53
## 1      C flood 0.68   9.63   bcd   9.59   9.68
## 5      D flood 0.50   9.48    cd   9.45   9.51
## 3      A flood 0.67   9.10    cd   9.06   9.15
## 9      E flood 0.99   7.88     d   7.82   7.95

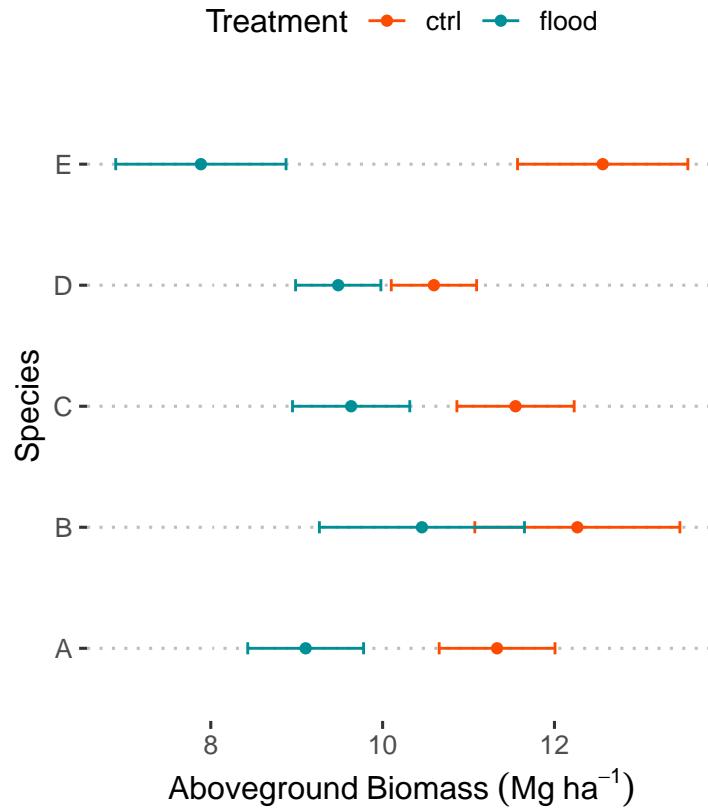
ggplot(mean_comparisons, aes(emmean, species))+
  geom_errorbarh(aes(xmin = emmean-SE, xmax = emmean + SE, color = trt), height = 0.1)+
  geom_point(aes(color = trt) , position = "dodge")+
  scale_fill_manual(values = c("#FC4C02", "#008E97"))+
  scale_color_manual(values = c("#FC4C02", "#008E97"))+
  theme_pubclean()+
  labs(x = expression(Aboveground~Biomass~(Mg~ha^{-1}))),
  y = "Species",

```

```

    color = "Treatment",
    fill = "Treatment")+
theme(aspect.ratio = 1)

```



```

plot_data <- expand.grid(doy = unique(dd_finalproj$doy),
                         species = c("A", "B", "C", "D", "E"),
                         trt = c("ctrl", "flood"))

plot_data <- bind_cols(plot_data,
                       predict(m3, newdata = plot_data, type = "link"))

## New names:
## * `` -> `...4`
## * `` -> `...5`

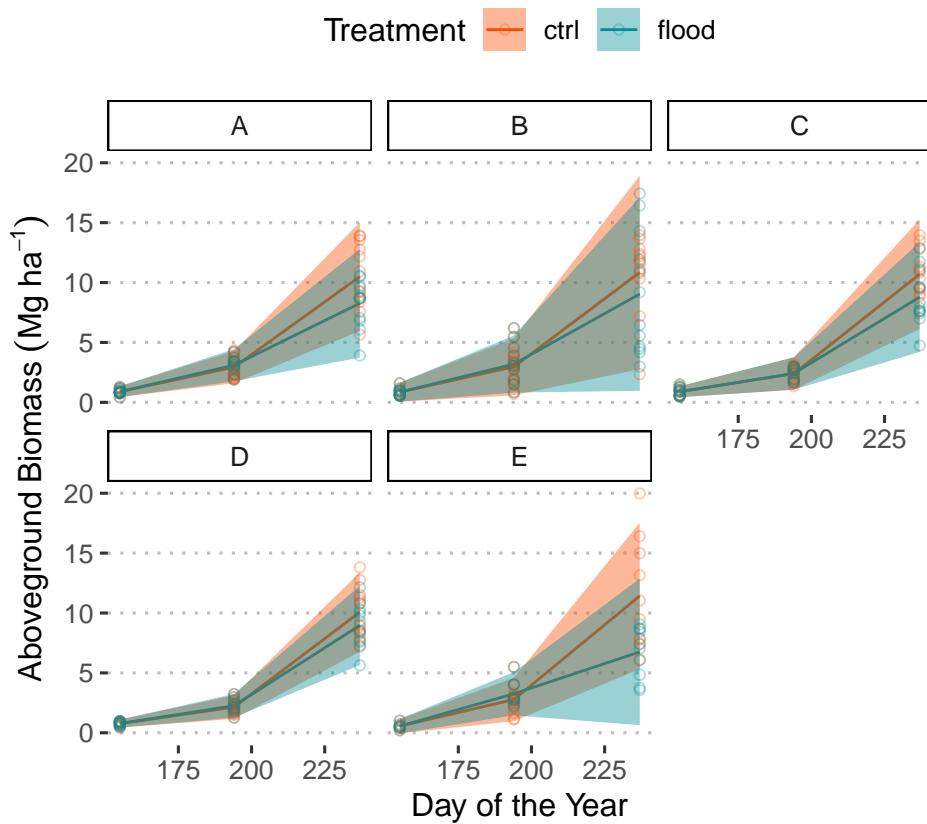
plot_data <- plot_data %>%
  rename(fitted = `...4`, sd = `...5`) %>%
  mutate(sd = .01+exp(sd))

plot_data %>%
  ggplot(aes(doy, fitted))+
  geom_line(aes(color = trt))+
  geom_ribbon(aes(ymin = fitted-sd*1.96, ymax = fitted+sd*1.96, fill = trt), alpha = .4)+
  geom_point(aes(y = agb_g, color = trt), data = dd_finalproj, shape = 21, alpha = .3)+
```

```

facet_wrap(~species)+
scale_fill_manual(values = c("#FC4C02", "#008E97"))+
scale_color_manual(values = c("#FC4C02", "#008E97"))+
labs(y = expression(Aboveground~Biomass~(Mg~ha^{-1})),
x= "Day of the Year",
color = "Treatment",
fill = "Treatment")+
theme_pubclean()+
theme(aspect.ratio = 1,
strip.background = element_rect(fill = NA, color = "black"))

```



Bonus: What if we hadn't changed our model?

This is just a demonstration of what happens when we design a model that does not describe our data generating process well. This does not necessarily have to go in your project.

```

plot_data <- expand.grid(doy = unique(dd_finalproj$doy),
                         species = c("A", "B", "C", "D", "E"),
                         trt = c("ctrl", "flood"))

plot_data <- bind_cols(plot_data,
                       predict(m1, newdata = plot_data, interval = "prediction"))

plot_data %>%

```

```

ggplot(aes(doy, fit))+  

  geom_line(aes(color = trt))+  

  geom_ribbon(aes(ymin = lwr, ymax = upr, fill = trt), alpha = .4)+  

  geom_point(aes(y = agb_g, color = trt), data = dd_finalproj, shape = 21, alpha = .3)+  

  facet_wrap(~species)+  

  scale_fill_manual(values = c("#FC4C02", "#008E97"))+  

  scale_color_manual(values = c("#FC4C02", "#008E97"))+  

  labs(y = expression(Aboveground~Biomass~(Mg~ha^{-1})),  

       x= "Day of the Year",  

       color = "Treatment",  

       fill = "Treatment") +  

  theme_pubclean() +  

  theme(aspect.ratio = 1,  

        strip.background = element_rect(fill = NA, color = "black"))

```

